Comparison of Blocking Agents for ELISA

Rachel Pearce Pratt and Bruce Roser, Quadrant Research Foundation, Cambridge Research Laboratories, Cambridge, England.

Introduction

The high sensitivity of ELISA implies a stringent limitation to the acceptable background signal due to non-specifically bound reactants. Low background is usually achieved by thorough "blocking" of the test wells with an inert or irrelevant protein.

This work demonstrates a variation in efficiency of blocking agents dependent on their molecular weight.

Methods and Results

Flat bottomed 96 well polystyrene plates (Thermo Scientific Nunc Immuno Plate MaxiSorp F96) were used in all experiments.

We compared the three blocking agents: bovine serum albumin (BSA), newborn calf serum (NBCS), and casein (i.e. sodium caseinate: 25 g casein powder (Sigma) dissolved in 800 mL 0.3 M NaOH by overnight stirring at 37°C, then titrated to pH 7.0 with HCI and made up to 1000 mL). After incubation of the wells with a dilution series in PBS of the respective blocking agents overnight at 4°C, and subsequent incubation with 1:4 PBS diluted rat serum for 90 min. at 4°C, each followed by washing with PBS + 0.05% Tween 20, the leakiness of the blocking layer for rat Ig was detected using the signal producing reagents: anti-rat rabbit F(ab')2 conjugated with horse radish peroxidase (HRP), and H₂O₂/tetramethyl-benzidine (TMB) enzyme substrate.

The results, which are shown in Fig. 1, confirm the superiority of casein for blocking agent.

In order to investigate the nature of the blocking leakiness, the two enzymes HRP and alkaline phosphatase (AP) were used. The wells were coated with one enzyme, blocked with NBCS or casein, and incubated with a dilution series of the other enzyme followed by addition to separate wells of the respective substrates: $\rm H_2O_2/TMB$ and para-nitrophenyl phosphate (PNPP).

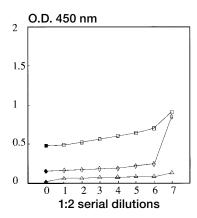


Fig. 1
Efficiency of three blocking agents.

Neither 100 mg/mL BSA (■) nor neat NBCS (♠) block the immobilization of rat Ig from 1:4 diluted serum as effectively as 25 mg/mL casein (♠). On dilution (open symbols) the failure of blocking is also more striking with the former agents than with casein.



Substrate for 1st stage enzyme was added to detect any displacement of that enzyme, whereas substrate for 2nd stage enzyme was added to detect penetration of the blocking layers by that other enzyme.

The results, which are shown in Fig. 2, indicate that blocking leakiness is rather due to penetration than to displacement of the blocking layers.

Again, attention was drawn to casein, specifically, why was it a better blocking agent?

HPLC gel filtration of casein and NBCS in PBS on a TSK HW 55S column (LKB, Sweden) showed that NBCS consists of protein species > 60 kD while casein contains a heterogeneous collection of molecular species from > 60 kD to < 10 kD (Fig. 3).

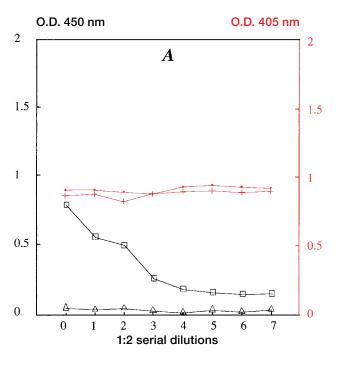
Fractionation of casein by ultrafiltration into three molecular weight (MW) grades, MW < 10 kD, MW > 30 kD, and 10 kD < MW < 30 kD, enabled us to show that the high blocking efficiency of casein was entirely due to its content of small MW proteins, and that the blocking potency was "inversely proportional" to the MW of the component proteins (Fig. 4).

Summary

Blocking leakiness seems to be due to penetration and not to displacement of blocking layers by 2nd stage reagents.

Casein appears to be an effective blocking agent due to its content of small protein species.

BSA and NBCS contain relatively large MW components so that random close packing of these molecules leaves bare patches of unblocked plastic surface to which 2nd stage reagents can bind, leading to higher background characteristics of these blocking agents.



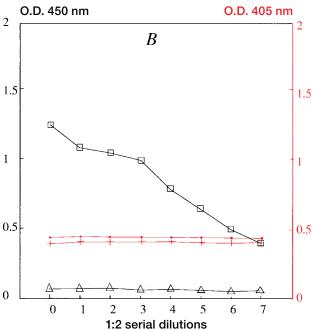


Fig. 2
Penetration of "blocking" AP by HRP (A), and vice versa (B).

A saturating layer of 1st stage enzyme (\blacksquare), supplemented with a secondary blocking with 10% v/v NBCS, was penetrated by 2nd stage enzyme in a dose-dependent manner (\square). When the secondary blocking was with 25 mg/mL casein, no penetration occurred at any dose of 2nd stage enzyme (\triangle). There was no significant displacement of 1st stage enzyme by the secondary reagents (+). Converted HRP and AP substrates were measured at 450 and 405 nm, respectively.

Gel filtration analysis of NBCS and sodium caseinate showing that whereas there are very few molecules smaller than 45 kD in NBCS, the size of the casein components is very heterogeneous with many molecules smaller than 6 kD. The MW standards refer to BSA (67 kD), ovalbumin (45 kD), a-lactalbumin (12.8 kD), and insulin (6 kD).

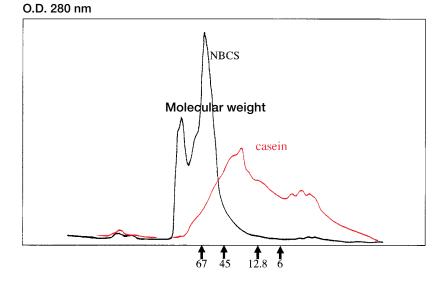
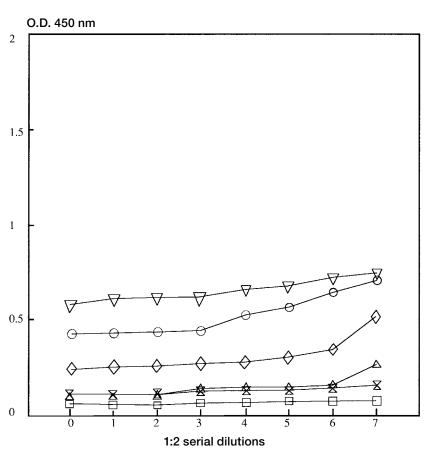


Fig. 4 Molecular size and blocking efficiency. Blocking of rat Ig immobilization with casein components from size fractionation confirms that smaller molecules block better. Rat Ig penetrated blocking layers of BSA (\bigtriangledown) and NBCS (\bigcirc) giving high plateau background levels and early failure of blocking with dilution. Casein molecules > 30 kD (♦) were much less effective than molecules < 10 kD (\square). Casein molecules between 10 and 30 kD were intermediate in effectiveness (\triangle), comparable with unfractionated casein (X). The initial concentration of each blocking agent corresponds to E280 = 4.0.



Further Reading

Batteiger B., Newhall V.W.J. & Jones R.B. (1987).

The use of Tween 20 as blocking agent in the immunological detection of proteins transferred to nitrocellulose membranes.

J. Immunol. Methods 55, 297-307.

Bird C.R., Gearing A.J.H. & Thorpe R. (1988). The use of Tween 20 alone as a blocking agent in Immunoblotting can cause artefactual results. J. Immunol. Methods 106, 175-179.

Engvall E. (1981). Methods in Enzymology 70, 419. Eds. Langone & Numakis. Acad. Press, London.

Fleming J.O. & Pen L.B. (1988).

Measurement of the concentration of murine IgG monoclonal antibody in hybridoma supernatant and ascites in absolute units by sensitive and reliable enzymelinked immunosorbent assays (ELISA).

J. Immunol. Methods 110, 11-18.

Gardas A. & Lewartowska A. (1988). Coating of proteins to polystyrene ELISA plates in the presence of detergents. J. Immunol. Methods 106, 251-255.

Hoffman W.L. & Jump A.A. (1986). Tween 20 removes antibodies and other proteins from nitrocellulose. J. Immunol. Methods 94, 191.

Pearce Pratt R. & Roser B. (1989). False positive signals in enzyme immunoassay (EIA): (II) Non antigen specific binding between Ig subclasses. Submitted.

Acknowledgements

This work was partly funded by MRC Programme grant No. PG8215534.

thermoscientific.com

© 2014 Thermo Fisher Scientific Inc. All rights reserved. All (other) trademarks are the property of Thermo Fisher Scientific Inc. and its subsidiaries.

ANZ: Australia: 1300 735 292, New Zealand: 0800 933 966; Asia: China Toll-free: 800-810-5118 or 400-650-5118; India: +91 22 6716 2200, India Toll-free: 1 800 22 8374; Japan: +81-3-5826-1616; Other Asian countries: 65 68729717 Europe: Austria: +43 1 801 40 0; Belgium: +32 2 482 30 30; Denmark: +45 4631 2000; France: +33 2 2803 2180; Germany: +49 6184 90 6000, Germany Toll-free: 0800 1-536 376; Italy: +39 02 95059 554; Netherlands: +31 76 571 4440; Nordic/Baltic countries: +358 9 329 10200; Russia/CIS: +7 (812) 703 42 15; Spain/Portugal: +34 93 223 09 18; Switzerland: +41 44 454 12 22; UK/Ireland: +44 870 609 9203 North America: USA/Canada +1 585 586 8800; USA Toll-free: 800 625 4327

South America: USA sales support: +1 585 899 7198 Countries not listed: +49 6184 90 6000 or +33 2 2803 2000

Thermo scientific